An Introduction to the Joint Modeling of Longitudinal and Survival Data, with Applications in R

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- *•* Often in follow-up studies different types of outcomes are collected
- *•* Explicit outcomes

◃ multiple longitudinal responses (e.g., markers, blood values)

◃ time-to-event(s) of particular interest (e.g., death, relapse)

• Implicit outcomes

◃ missing data (e.g., dropout, intermittent missingness)

◃ random visit times

- *•* Methods for the separate analysis of such outcomes are well established in the literature
- *•* Survival data:
	- *◃* Cox model, accelerated failure time models, . . .
- *•* Longitudinal data
	- *◃* mixed effects models, GEE, marginal models, . . .

Purpose of this course is to present the state of the art in

Joint Modeling Techniques for Longitudinal and Survival Data

- *•* **Goals:** After this course participants will be able to
	- \triangleright identify settings in which a joint modeling approach is required,
	- *◃* construct and fit an appropriate joint model, and
	- *◃* correctly interpret the obtained results
- *•* The course will be explanatory rather than mathematically rigorous
	- *◃* emphasis is given on sufficient detail in order for participants to obtain a clear view on the different joint modeling approaches, and how they should be used in practice

• **Part I:** Introduction

- *◃* Data sets that we will use throughout the course
- *◃* Categorization of possible research questions
- *•* **Part II:** (brief) Review of Linear Mixed Models
	- *◃* Features of repeated measurements data
	- *◃* Linear mixed models
	- *◃* Missing data in longitudinal studies

- *•* **Part III:** (brief) Review of Relative Risk Models
	- *◃* Features of survival data
	- *◃* Relative risk models
	- *◃* Time-dependent covariates
- *•* **Part IV:** The Basic Joint Model
	- *◃* Definition
	- *◃* Estimation & Inference
	- *◃* Connection with the missing data framework

- *•* **Part V:** Extensions of the Basic Joint Model
	- *◃* Parameterizations
	- *◃* Latent class joint models
	- *◃* Other extensions for the longitudinal and survival submodels (briefly)

• **Part VI:** Dynamic Predictions

- \rhd Individualized predictions for the survival
- \rhd Effect of the parameterization

- *•* Lectures & short software practicals using R package **JM** and/or **JMbayes**
- *•* Material:
	- *◃* Course Notes
	- **⊳** R code in soft format
- Within the course notes there are several examples of R code which are denoted by the symbol 'R> '

- *•* Joint modeling sources*[∗]*
	- *◃* Rizopoulos, D. (2012). *Joint Models for Longitudinal and Time-to-Event Data, with Applications in R*. Boca Raton: Chapman & Hall/CRC.
	- *◃* Fitzmaurice, G., Davidian, M., Verbeke, G. and Molenberghs, G. (2009). *Longitudinal Data Analysis*. Handbooks of Modern Statistical Methods. Boca Raton: Chapman & Hall/CRC, Chapter 15.
	- *◃* Wu, L. (2009). *Mixed Effects Models for Complex Data*. Boca Raton: Chapman & Hall/CRC, Chapter 8.
	- *◃* Ibrahim, J., Chen, M.-H. and Sinha, D. (2001). *Bayesian Survival Analysis*. New York: Springer-Verlag, Chapter 7.

∗ extra references of papers using joint modeling available at pp. 175–182.

- *•* Useful material for package **JM** can be found in the web sites:
	- *◃* http://jmr.r-forge.r-project.org [R code used in the book]
	- *◃* https://eur.academia.edu/DimitrisRizopoulos/Teaching-Documents [additional R script files]
- *•* Useful material for package **JMbayes**

 ρ a paper describing the current capabilities of the package is available on arXiv http://arxiv.org/abs/1404.7625

• Blog about joint modeling http://iprogn.blogspot.nl/

- *•* Other software packages capable of fitting joint models
	- *◃* in **R**: **joineR** (by Philipson et al.), **lcmm** (by Proust-Lima et al.) *◃* in **SAS**: **%JM** macro (by Garcia-Hernandez and Rizopoulos – http://www.jm-macro.com/), **%JMFit** macro (by Zhang et al.) *◃* in **STATA**: **stjm** (by Crowther)

- *•* Standard texts in survival analysis
	- *◃* Kalbfleisch, J. and Prentice, R. (2002). *The Statistical Analysis of Failure Time Data, 2nd Ed.*. New York: Wiley.
	- *◃* Therneau, T. and Grambsch, P. (2000). *Modeling Survival Data: Extending the Cox Model*. New York: Springer-Verlag.
	- *◃* Cox, D. and Oakes, D. (1984). *Analysis of Survival Data*. London: Chapman & Hall.
	- *◃* Andersen, P., Borgan, O., Gill, R. and Keiding, N. (1993). *Statistical Models Based on Counting Processes*. New York: Springer-Verlag.
	- *◃* Klein, J. and Moeschberger, M. (2003). *Survival Analysis Techniques for Censored and Truncated Data*. New York: Springer-Verlag.

- *•* Standard texts in longitudinal data analysis
	- *◃* Verbeke, G. and Molenberghs, G. (2000). *Linear Mixed Models for Longitudinal Data*. New York: Springer-Verlag.
	- *◃* Molenberghs, G. and Verbeke, G. (2005). *Models for Discrete Longitudinal Data*. New York: Springer-Verlag.
	- *◃* Fitzmaurice, G., Laird, N., and Ware, J. (2004). *Applied Longitudinal Analysis*. Hoboken: Wiley.
	- *◃* Diggle, P., Heagerty, P., Liang, K.-Y., and Zeger, S. (2002). *Analysis of Longitudinal Data*, 2nd edition. New York: Oxford University Press.

Chapter 1 Introduction

- AIDS: 467 HIV infected patients who had failed or were intolerant to zidovudine therapy (AZT) (Abrams et al., NEJM, 1994)
- *•* The aim of this study was to compare the efficacy and safety of two alternative antiretroviral drugs, didanosine (ddI) and zalcitabine (ddC)
- *•* Outcomes of interest:
	- **⊳** time to death
	- *◃* randomized treatment: 230 patients ddI and 237 ddC
	- *◃* CD4 cell count measurements at baseline, 2, 6, 12 and 18 months
	- *◃* prevOI: previous opportunistic infections

1.1 Motivating Longitudinal Studies (cont'd)

Kaplan−Meier Estimate

• Research Questions:

- *◃* How strong is the association between CD4 cell count and the risk for death?
- *◃* Is CD4 cell count a good biomarker?
	- * if treatment improves CD4 cell count, does it also improve survival?

- **PBC:** Primary Biliary Cirrhosis:
	- *◃* a chronic, fatal but rare liver disease
	- *◃* characterized by inflammatory destruction of the small bile ducts within the liver
- Data collected by Mayo Clinic from 1974 to 1984 (Murtaugh et al., Hepatology, 1994)
- *•* Outcomes of interest:
	- \triangleright time to death and/or time to liver transplantation
	- *◃* randomized treatment: 158 patients received D-penicillamine and 154 placebo
	- *◃* longitudinal serum bilirubin levels

1.1 Motivating Longitudinal Studies (cont'd)

Kaplan−Meier Estimate

• Research Questions:

- *◃* How strong is the association between bilirubin and the risk for death?
- *◃* How the observed serum bilirubin levels could be utilized to provide predictions of survival probabilities?
- *◃* Can bilirubin discriminate between patients of low and high risk?

- Depending on the questions of interest, different types of statistical analysis are required
- *•* We will distinguish between two general types of analysis
	- *◃* separate analysis per outcome
	- *◃* joint analysis of outcomes
- *•* Focus on each outcome separately
	- *◃* does treatment affect survival?
	- *◃* are the average longitudinal evolutions different between males and females?

 \triangleright ...

- *•* Focus on multiple outcomes
	- *◃* Complex hypothesis testing: does treatment improve the average longitudinal profiles in all markers?
	- *◃* Complex effect estimation: how strong is the association between the longitudinal evolution of CD4 cell counts and the hazard rate for death?
	- *◃* Association structure among outcomes:
		- * how the association between markers evolves over time (evolution of the association)
		- * how marker-specific evolutions are related to each other (association of the evolutions)

◃ Prediction: can we improve prediction for the time to death by considering all markers simultaneously?

◃ Handling implicit outcomes: focus on a single longitudinal outcome but with dropout or random visit times

- *•* Up to now emphasis has been
	- *◃ restricted* or *coerced* to separate analysis per outcome
	- *◃* or given to naive types of joint analysis (e.g., last observation carried forward)
- *•* Main reasons
	- *◃* lack of appropriate statistical methodology
	- *◃* lack of efficient computational approaches & software

- *•* However, recently there has been an explosion in the statistics and biostatistics literature of joint modeling approaches
- *•* Many different approaches have been proposed that
	- *◃* can handle different types of outcomes
	- \rhd can be utilized in pragmatic computing time
	- *◃* can be rather flexible
	- *◃* **most importantly:** can answer the questions of interest

- Let Y_1 and Y_2 two outcomes of interest measured on a number of subjects for which joint modeling is of scientific interest
	- *◃* both can be measured longitudinally
	- *◃* one longitudinal and one survival
- We have various possible approaches to construct a joint density $p(y_1, y_2)$ of $\{Y_1, Y_2\}$ \triangleright Conditional models: $p(y_1, y_2) = p(y_1)p(y_2 \mid y_1)$
	- $\rho \in \text{Copulas: } p(y_1, y_2) = c\{\mathcal{F}(y_1), \mathcal{F}(y_2)\} p(y_1) p(y_2)$

But **Random Effects Models** have (more or less) prevailed

• Random Effects Models specify

$$
p(y_1, y_2) = \int p(y_1, y_2 | b) p(b) db
$$

=
$$
\int p(y_1 | b) p(y_2 | b) p(b) db
$$

 \triangleright Unobserved random effects b explain the association between Y_1 and Y_2

◃ Conditional Independence assumption

$$
Y_1 \perp \!\!\! \perp Y_2 \mid b \mid
$$

• Features:

 $P \triangleright Y_1$ and Y_2 can be of different type * one continuous and one categorical * one continuous and one survival $*$. . .

◃ Extensions to more than two outcomes straightforward

 \triangleright Specific association structure between Y_1 and Y_2 is assumed

◃ Computationally intensive (especially in high dimensions)
Chapter 2

Linear Mixed-Effects Models

- *•* Repeated evaluations of the same outcome in each subject in time
	- *◃* CD4 cell count in HIV-infected patients
	- *◃* serum bilirubin in PBC patients
- *•* Longitudinal studies allow to investigate
	- 1. how treatment means differ at specific time points, e.g., at the end of the study (*cross-sectional effect*)
	- 2. how treatment means or differences between means of treatments change over time (*longitudinal effect*)

Measurements on the same subject are expected to be (positively) correlated

• This implies that standard statistical tools, such as the *t*-test and simple linear regression that assume independent observations, are not optimal for longitudinal data analysis.

• The direct approach to model correlated data *⇒ multivariate regression*

$$
y_i = X_i \beta + \varepsilon_i, \quad \varepsilon_i \sim \mathcal{N}(0, V_i),
$$

where

 \triangleright y_i the vector of responses for the *i*th subject

- $\triangleright X_i$ design matrix describing structural component
- $\triangleright V_i$ covariance matrix describing the correlation structure
- *•* There are several options for modeling *Vⁱ* , e.g., compound symmetry, autoregressive process, exponential spatial correlation, Gaussian spatial correlation, . . .

• **Alternative intuitive approach:** Each subject in the population has her own subject-specific mean response profile over time

• The evolution of each subject in time can be described by a linear model

$$
y_{ij} = \tilde{\beta}_{i0} + \tilde{\beta}_{i1} t_{ij} + \varepsilon_{ij}, \quad \varepsilon_{ij} \sim \mathcal{N}(0, \sigma^2),
$$

where

 $\triangleright y_{ij}$ the *j*th response of the *i*th subject

 $\rhd \widetilde{\beta}_{i0}$ is the intercept and $\widetilde{\beta}_{i1}$ the slope for subject i

• **Assumption:** Subjects are randomly sampled from a population *⇒* subject-specific regression coefficients are also sampled from a population of regression coefficients

$$
\tilde{\beta}_i \thicksim \mathcal{N}(\beta, D)
$$

• We can reformulate the model as

$$
y_{ij} = (\beta_0 + b_{i0}) + (\beta_1 + b_{i1})t_{ij} + \varepsilon_{ij},
$$

where

*◃ β*s are known as the *fixed effects* \triangleright b_i s are known as the *random effects*

• In accordance for the random effects we assume

$$
b_i = \begin{bmatrix} b_{i0} \\ b_{i1} \end{bmatrix} \sim \mathcal{N}(0, D)
$$

• Put in a general form

$$
\begin{cases}\ny_i = X_i \beta + Z_i b_i + \varepsilon_i, \\
b_i \sim \mathcal{N}(0, D), \quad \varepsilon_i \sim \mathcal{N}(0, \sigma^2 I_{n_i}),\n\end{cases}
$$

with

◃ X design matrix for the fixed effects *β* \triangleright *Z* design matrix for the random effects b_i $\triangleright b_i \perp \!\!\! \perp \varepsilon_i$

- *•* Interpretation:
	- $\rho \triangleright \beta_j$ denotes the change in the average y_i when x_j is increased by one unit
	- \triangleright \dot{b}_i are interpreted in terms of how a subset of the regression parameters for the *i*th subject deviates from those in the population
- Advantageous feature: population $+$ subject-specific predictions
	- β describes mean response changes in the population
	- $\beta + b_i$ describes individual response trajectories

• Example: We fit a linear mixed model for the AIDS dataset assuming *◃* different average longitudinal evolutions per treatment group (fixed part) **▷ random intercepts & random slopes (random part)**

$$
\begin{cases}\ny_{ij} = \beta_0 + \beta_1 t_{ij} + \beta_2 \{\text{ddI}_i \times t_{ij}\} + b_{i0} + b_{i1} t_{ij} + \varepsilon_{ij}, \\
b_i \sim \mathcal{N}(0, D), \quad \varepsilon_{ij} \sim \mathcal{N}(0, \sigma^2)\n\end{cases}
$$

• Note: We did not include a main effect for treatment due to randomization

• No evidence of differences in the average longitudinal evolutions between the two treatments

- R> There are two primary packages in R for mixed models analysis:
	- *◃* Package **nlme**
		- * fits linear & nonlinear mixed effects models, and marginal models for normal data
		- * allows for both random effects & correlated error terms
		- * several options for covariances matrices and variance functions
	- *◃* Package **lme4**
		- * fits linear, nonlinear & generalized mixed effects models
		- * uses only random effects
		- * allows for nested and crossed random-effects designs

- R> We will only use package **nlme** because package **JM** accepts as an argument a linear mixed model fitted by **nlme**
- R> The basic function to fit linear mixed models is $\text{Im}(\epsilon)$ and has three basic arguments *◃* fixed: a formula specifying the response vector and the fixed-effects structure *◃* random: a formula specifying the random-effects structure **▷ data:** a data frame containing all the variables

R> The data frame that contains all variables should be in the *long format*

R> Using formulas in R

$$
\triangleright \text{CD4} = \text{Time} + \text{Gender} \n\Rightarrow \boxed{\text{cd4}} \sim \text{time} + \text{gender}
$$

$$
\triangleright \text{CD4} = \text{Time} + \text{Gender} + \text{Time*Gender}
$$
\n
$$
\Rightarrow \text{Cd4} \sim \text{time} + \text{gender} + \text{time:gender}
$$
\n
$$
\Rightarrow \text{Cd4} \sim \text{time*gender} \text{ (the same)}
$$

$$
\triangleright \text{CD4} = \text{Time} + \text{Time}^2
$$

$$
\Rightarrow \boxed{\text{cd4}} \sim \text{time} + \text{I}(\text{time}^2)
$$

R> Note: the intercept term is included by default

R> The code used to fit the linear mixed model for the AIDS dataset (p. 28) is as follows

```
lmeFit \leq 1me(CD4 \sim obstime + obstime:drug, data = aids,
    random = \sim obstime | patient)
```

```
summary(lmeFit)
```


R> The same fixed-effects structure but only random intercepts

```
lme(CD4 ~ obstime + obstime:drug, data = aids,
    random = \degree 1 | patient)
```
R> The same fixed-effects structure, random intercepts & random slopes, with a diagonal covariance matrix (using the pdDiag() function)

```
lme(CD4 ~ obstime + obstime:drug, data = aids,
   random = list(patient = pdDiag(form = ~' obstime)))
```


- A major challenge for the analysis of longitudinal data is the problem of missing data
	- *◃* studies are designed to collect data on every subject at a set of prespecified follow-up times
	- *◃* often subjects miss some of their planned measurements for a variety of reasons
- *•* We can have different patterns of missing data

◃ Subject 1: Completer

◃ Subject 2: dropout

◃ Subject 3: late entry

◃ Subject 4: intermittent

• Implications of missingness:

◃ we collect less data than originally planned *⇒ loss of efficiency*

◃ not all subjects have the same number of measurements *⇒ unbalanced datasets*

- *◃* missingness may depend on outcome *⇒ potential bias*
- *•* For the handling of missing data, we introduce the missing data indicator

$$
r_{ij} = \left\{ \begin{array}{ll} 1 \ \ \text{if} \ y_{ij} \ \text{is observed} \\ 0 \ \ \text{otherwise} \end{array} \right.
$$

• We obtain a partition of the complete response vector *yⁱ*

 \triangleright observed data y_i^o i , containing those y_{ij} for which $r_{ij} = 1$

 \triangleright missing data y_i^m i^{m}_{i} , containing those y_{ij} for which $r_{ij}=0$

• **For the remaining we will focus on dropout** *⇒* notation can be simplified

$$
\triangleright \text{ Discrete dropout time: } r_i^d = 1 + \sum_{j=1}^{n_i} r_{ij} \text{ (ordinal variable)}
$$

◃ **Continuous time**: *T ∗* \mathbf{r}_i^* denotes the time to dropout

- To describe the probabilistic relation between the measurement and missingness processes Rubin (1976, Biometrika) has introduced three mechanisms
- *• Missing Completely At Random (MCAR)*: The probability that responses are missing is unrelated to both y_i^o $\frac{a}{i}$ and y_i^m *i*

$$
p(r_i \mid y_i^o, y_i^m) = p(r_i)
$$

• Examples

- *◃* subjects go out of the study after providing a pre-determined number of measurements
- *◃* laboratory measurements are lost due to equipment malfunction

- *•* Features of MCAR:
	- \triangleright The observed data y_i^o $_i^o$ can be considered a random sample of the complete data y_i
	- *◃* We can use any statistical procedure that is valid for complete data
		- * sample averages per time point
		- * linear regression, ignoring the correlation (consistent, but not efficient)
		- * *t*-test at the last time point
		- $*$. . .

• Missing At Random (MAR): The probability that responses are missing is related to y_i^o $\frac{o}{i}$, but is unrelated to y_i^m *i*

$$
p(r_i \mid y_i^o, y_i^m) = p(r_i \mid y_i^o)
$$

• Examples

◃ study protocol requires patients whose response value exceeds a threshold to be removed from the study

◃ physicians give rescue medication to patients who do not respond to treatment

- *•* Features of MAR:
	- *◃* The observed data cannot be considered a random sample from the target population
	- *◃* Not all statistical procedures provide valid results

• Missing Not At Random (MNAR): The probability that responses are missing is related to y_i^m $\frac{m}{i}$, and possibly also to y_i^o *i*

$$
p(r_i \mid y_i^m) \quad \text{or} \quad p(r_i \mid y_i^o, y_i^m)
$$

• Examples

◃ in studies on drug addicts, people who return to drugs are less likely than others to report their status

◃ in longitudinal studies for quality-of-life, patients may fail to complete the questionnaire at occasions when their quality-of-life is compromised

- *•* Features of MNAR
	- *◃* The observed data cannot be considered a random sample from the target population
	- \triangleright Only procedures that explicitly model the joint distribution $\{y^o_i\}$ $\left\{ \begin{array}{c} c_i, y_i^m, r_i \end{array} \right\}$ provide valid inferences *⇒* **analyses which are valid under MAR will not be valid under MNAR**

We cannot tell from the data at hand whether the missing data mechanism is MAR or MNAR

Note: We can distinguish between MCAR and MAR

Chapter 3 Relative Risk Models

- *•* The most important characteristic that distinguishes the analysis of time-to-event outcomes from other areas in statistics is **Censoring**
	- *◃* the event time of interest is not fully observed for all subjects under study
- Implications of censoring:
	- \triangleright standard tools, such as the sample average, the t -test, and linear regression **cannot** be used
	- *◃* inferences may be sensitive to misspecification of the distribution of the event times

- Several types of censoring:
	- *◃* Location of the true event time wrt the censoring time: *right*, *left* & *interval*
	- *◃* Probabilistic relation between the true event time & the censoring time: *informative* & *non-informative* (similar to MNAR and MAR)

Here we focus on non-informative right censoring

• Note: Survival times may often be truncated; analysis of truncated samples requires similar calculations as censoring

- *•* Notation (*i* denotes the subject)
	- $\triangleright T_i^*$ i^* 'true' time-to-event
	- $\triangleright C_i$ the censoring time (e.g., the end of the study or a random censoring time)
- *•* Available data for each subject
	- \triangleright observed event time: $T_i = \min(T_i^*)$ $C_i^*, C_i)$
	- \rhd event indicator: $\delta_i = 1$ if event; $\delta_i = 0$ if censored

Our aim is to make valid inferences for *T ∗ ⁱ* **but using** $\textbf{only} \ \{T_i, \delta_i\}$

• **Relative Risk Models** assume a multiplicative effect of covariates on the hazard scale, i.e.,

$$
h_i(t) = h_0(t) \exp(\gamma_1 w_{i1} + \gamma_2 w_{i2} + \ldots + \gamma_p w_{ip}) \Rightarrow
$$

$$
\log h_i(t) = \log h_0(t) + \gamma_1 w_{i1} + \gamma_2 w_{i2} + \ldots + \gamma_p w_{ip},
$$

where

 $\triangleright h_i(t)$ denotes the hazard for an event for patient *i* at time *t* $\rhd h_0(t)$ denotes the baseline hazard \triangleright w_{i1}, \ldots, w_{ip} a set of covariates

• Standard MLE can be applied based on the log-likelihood function

$$
\ell(\theta) = \sum_{i=1}^n \delta_i \log p(T_i; \theta) + (1 - \delta_i) \log S_i(T_i; \theta),
$$

which also can be re-expressed in terms of the hazard function

$$
\ell(\theta) = \sum_{i=1}^{n} \delta_i \log h_i(T_i; \theta) - \int_0^{T_i} h_i(s; \theta) ds
$$

Sensitivity to distributional assumptions due to censoring

- *•* **Cox Model:** We make no assumptions for the baseline hazard function
- *•* Parameter estimates and standard errors are based on the log partial likelihood function

$$
p\ell(\gamma) = \sum_{i=1}^n \delta_i \Big[\gamma^\top w_i - \log \Big\{ \sum_{j: T_j \ge T_i} \exp(\gamma^\top w_j) \Big\} \Big],
$$

where only patients who had an event contribute

• Example: For the PBC dataset were interested in the treatment effect while correcting for sex and age effects

$$
h_i(t) \ = \ h_0(t) \exp(\gamma_1 \texttt{D-penic}_i + \gamma_2 \texttt{Female}_i + \gamma_3 \texttt{Age}_i)
$$

- R> The primary package in R for the analysis of survival data is the **survival** package
- R> A key function in this package that is used to specify the available event time information in a sample at hand is Surv()
- R> For right censored failure times (i.e., what we will see in this course) we need to provide the observed event times time, and the event indicator status, which equals 1 for true failure times and 0 for right censored times

Surv(time, status)

R> Cox models are fitted using function coxph(). For instance, for the PBC data the following code fits the Cox model that contains the main effects of 'drug', 'sex' and 'age':

```
coxFit \leq coxph(Surv(years, status2) \sim drug + sex + age,
data = pbc2.id)
```

```
summary(coxFit)
```
R> The two main arguments are a formula specifying the design matrix of the model and a data frame containing all the variables

- *•* Often interest in the association between a time-dependent covariate and the risk for an event
	- *◃* treatment changes with time (e.g., dose)
	- \rhd time-dependent exposure (e.g., smoking, diet)
	- *◃* markers of disease or patient condition (e.g., blood pressure, PSA levels)
	- \triangleright \ldots
- Example: In the PBC study, are the longitudinal bilirubin measurements associated with the hazard for death?

- *•* To answer our questions of interest we need to postulate a model that relates
	- *◃* the serum bilirubin with
	- *◃* the time-to-death
- *•* The association between **baseline** marker levels and the risk for death can be estimated with standard statistical tools (e.g., Cox regression)
- *•* When we move to the time-dependent setting, a more **careful consideration** is required

• There are two types of time-dependent covariates

```
(Kalbfleisch and Prentice, 2002, Section 6.3)
```
 \triangleright Exogenous (aka external): the future path of the covariate up to any time $t > s$ is not affected by the occurrence of an event at time point *s*, i.e.,

 $\Pr\{\mathcal{Y}_i(t) | \mathcal{Y}_i(s), T_i^* \geq s\} = \Pr\{\mathcal{Y}_i(t) | \mathcal{Y}_i(s), T_i^* = s\},$

where
$$
0 < s \leq t
$$
 and $\mathcal{Y}_i(t) = \{y_i(s), 0 \leq s < t\}$

◃ Endogenous (aka internal): not Exogenous

- *•* It is **very important** to distinguish between these two types of time-dependent covariates, because the type of covariate dictates the appropriate type of analysis
- *•* In our motivating examples all time-varying covariates are **Biomarkers** *⇒* These are always **endogenous** covariates
	- *◃* measured with error (i.e., biological variation)
	- \rhd the complete history is not available
	- *◃* existence directly related to failure status

3.4 Time Dependent Covariates (cont'd)

Subject 127

Erasmus MC

zafus

• The Cox model presented earlier can be extended to handle time-dependent covariates using the counting process formulation

$$
h_i(t | \mathcal{Y}_i(t), w_i) = h_0(t) R_i(t) \exp{\{\gamma^\top w_i + \alpha y_i(t)\}},
$$

where

- \triangleright $N_i(t)$ is a counting process which counts the number of events for subject *i* by time *t*,
- $\triangleright h_i(t)$ denotes the intensity process for $N_i(t)$,
- $\rhd R_i(t)$ denotes the at risk process ('1' if subject *i* still at risk at *t*), and
- \triangleright $y_i(t)$ denotes the value of the time-varying covariate at *t*

• Interpretation:

$$
h_i(t | \mathcal{Y}_i(t), w_i) = h_0(t) R_i(t) \exp{\{\gamma^\top w_i + \alpha y_i(t)\}}
$$

 $\exp(\alpha)$ denotes the relative increase in the risk for an event at time *t* that results from one unit increase in $y_i(t)$ at the same time point

• Parameters are estimated based on the log-partial likelihood function

$$
p\ell(\gamma,\alpha) = \sum_{i=1}^n \int_0^\infty \left\{ R_i(t) \exp\{\gamma^\top w_i + \alpha y_i(t)\} - \log \left[\sum_j R_j(t) \exp\{\gamma^\top w_j + \alpha y_j(t)\}\right] \right\} dN_i(t)
$$

• Typically, data must be organized in the long format

- *•* How does the extended Cox model handle time-varying covariates?
	- *◃* assumes no measurement error
	- *◃* step-function path
	- *◃* existence of the covariate is not related to failure status

• Therefore, the extended Cox model is only valid for exogenous time-dependent covariates

> **Treating endogenous covariates as exogenous may produce spurious results!**

Chapter 4 The Basic Joint Model

• To account for the special features of endogenous covariates a new class of models has been developed

Joint Models for Longitudinal and Time-to-Event Data

- Intuitive idea behind these models
	- 1. use an appropriate model to describe the evolution of the marker in time for each patient
	- 2. the estimated evolutions are then used in a Cox model
- *•* Feature: Marker level's are **not** assumed constant between visits

• Some notation

- $\triangleright T_i^*$ i^*_i : True event time for patient i
- $\triangleright T_i$: Observed event time for patient i
- $\rhd \delta_i$: Event indicator, i.e., equals 1 for true events
- $\triangleright y_i$: Longitudinal responses
- *•* We will formulate the joint model in 3 steps in particular, . . .

- Step 1: Let's assume that we know $m_i(t)$, i.e., the *true* & *unobserved* value of the marker at time *t*
- *•* Then, we can define a standard relative risk model

$$
h_i(t | \mathcal{M}_i(t)) = h_0(t) \exp{\{\gamma^\top w_i + \alpha m_i(t)\}},
$$

- *◃ Mi*(*t*) = *{mi*(*s*)*,* 0 *≤ s < t}* longitudinal history
- $\triangleright \alpha$ quantifies the strength of the association between the marker and the risk for an event
- \triangleright w_i baseline covariates

- Step 2: From the observed longitudinal response $y_i(t)$ reconstruct the covariate history for each subject
- *•* Mixed effects model (we focus, for now, on continuous markers)

$$
y_i(t) = m_i(t) + \varepsilon_i(t)
$$

= $x_i^{\top}(t)\beta + z_i^{\top}(t)b_i + \varepsilon_i(t), \quad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2),$

 $\triangleright x_i(t)$ and β : Fixed-effects part $\rho \approx z_i(t)$ and b_i : Random-effects part, $b_i \thicksim \mathcal{N}(0,D)$

- *•* Step 3: The two processes are associated *⇒* define a model for their joint distribution
- *•* Joint Models for such joint distributions are of the following form (Tsiatis & Davidian, Stat. Sinica, 2004)

$$
p(y_i, T_i, \delta_i) \ = \ \int p(y_i \mid b_i) \ \{ h(T_i \mid b_i)^{\delta_i} \ S(T_i \mid b_i) \} \ p(b_i) \ db_i,
$$

 $\triangleright b_i$ a vector of random effects that explains the interdependencies $p(\cdot)$ density function; $S(\cdot)$ survival function

- *•* Key assumption: **Full Conditional Independence** *⇒* random effects explain all interdependencies
	- *◃* the longitudinal outcome is independent of the time-to-event outcome
	- *◃* the repeated measurements in the longitudinal outcome are independent of each other

$$
p(y_i, T_i, \delta_i | b_i) = p(y_i | b_i) p(T_i, \delta_i | b_i)
$$

$$
p(y_i | b_i) = \prod_j p(y_{ij} | b_i)
$$

Caveat: CI is difficult to be tested

- *•* The censoring and visiting*[∗]* processes are assumed non-informative:
- *•* Decision to withdraw from the study or appear for the next visit
	- \triangleright **may depend** on observed past history (baseline covariates $+$ observed longitudinal responses)
	- *◃* **no additional dependence** on underlying, latent subject characteristics associated with prognosis

*[∗]*The visiting process is defined as the mechanism (stochastic or deterministic) that generates the time points at which longitudinal measurements are collected.

• The survival function, which is a part of the likelihood of the model, depends on the whole longitudinal history

$$
S_i(t | b_i) = \exp\left(-\int_0^t h_0(s) \exp\{\gamma^\top w_i + \alpha m_i(s)\} ds\right)
$$

- *•* Therefore, care in the definition of the design matrices of the mixed model *◃* when subjects have nonlinear profiles *⇒*
	- \rhd use splines or polynomials to model them flexibly

• Assumptions for the baseline hazard function $h_0(t)$

◃ parametric *⇒* possibly restrictive

◃ unspecified *⇒* within JM framework underestimates standard errors

• It is advisable to use parametric but flexible models for $h_0(t)$

◃ splines

$$
\log h_0(t) = \gamma_{h_0,0} + \sum_{q=1}^{Q} \gamma_{h_0,q} B_q(t,\nu),
$$

where

- * $B_q(t, v)$ denotes the q-th basis function of a B-spline with knots v_1, \ldots, v_Q
- * γ_{h_0} a vector of spline coefficients

- It is advisable to use parametric but flexible models for $h_0(t)$
	- *◃* step-functions: piecewise-constant baseline hazard often works satisfactorily

$$
h_0(t) = \sum_{q=1}^{Q} \xi_q I(v_{q-1} < t \le v_q),
$$

where $0 = v_0 < v_1 < \cdots < v_Q$ denotes a split of the time scale

- *•* Mainly maximum likelihood but also Bayesian approaches
- *•* The log-likelihood contribution for subject *i*:

$$
\ell_i(\theta) = \log \int \left\{ \prod_{j=1}^{n_i} p(y_{ij} \mid b_i; \theta) \right\} \left\{ h(T_i \mid b_i; \theta)^{\delta_i} S_i(T_i \mid b_i; \theta) \right\} p(b_i; \theta) db_i,
$$

$$
S_i(t | b_i; \theta) = \exp\left(-\int_0^t h_0(s; \theta) \exp\{\gamma^\top w_i + \alpha m_i(s)\} ds\right)
$$

- *•* Both integrals do not have, in general, a closed-form solution *⇒* need to be approximated numerically
- *•* Standard numerical integration algorithms
	- *◃* Gaussian quadrature
	- *◃* Monte Carlo
	- \triangleright . . .
- *•* More difficult is the integral with respect to *bⁱ* because it can be of high dimension
	- *◃* Laplace approximations
	- *◃* pseudo-adaptive Gaussian quadrature rules

• To maximize the approximated log-likelihood

$$
\ell(\theta) = \sum_{i=1}^n \log \int p(y_i | b_i; \theta) \left\{ h(T_i | b_i; \theta)^{\delta_i} S_i(T_i | b_i; \theta) \right\} p(b_i; \theta) db_i,
$$

we need to employ an optimization algorithm

- *•* Standard choices
	- \triangleright EM (treating b_i as missing data)
	- *◃* Newton-type
	- *◃* hybrids (start with EM and continue with quasi-Newton)

• Standard errors: Standard asymptotic MLE

$$
\text{var}(\hat{\theta}) \ = \ \left\{ - \sum_{i=1}^n \frac{\partial^2 \log p(y_i, T_i, \delta_i; \theta)}{\partial \theta^\top \partial \theta} \Big|_{\theta = \hat{\theta}} \right\}^{-1}
$$

- Standard asymptotic tests + information criteria
	- *◃* likelihood ratio test
	- *◃* score test
	- *◃* Wald test
	- **⊳** AIC, BIC, ...

• Based on a fitted joint model, estimates for the random effects are based on the posterior distribution:

$$
p(b_i | T_i, \delta_i, y_i; \theta) = \frac{p(T_i, \delta_i | b_i; \theta) p(y_i | b_i; \theta) p(b_i; \theta)}{p(T_i, \delta_i, y_i; \theta)}
$$

 $\propto p(T_i, \delta_i \mid b_i; \theta) p(y_i \mid b_i; \theta) p(b_i; \theta),$

in which θ is replaced by its MLE $\hat{\theta}$

• Measures of location

$$
\begin{cases} \bar{b}_i = \int b_i \, p(b_i \mid T_i, \delta_i, y_i; \hat{\theta}) \, db_i \\ \hat{b}_i = \operatorname{argmax}_{b} \{ \log p(b \mid T_i, \delta_i, y_i; \hat{\theta}) \} \end{cases}
$$

• Measures of dispersion

$$
\left\{\begin{aligned} \text{var}(b_i) &= \int (b_i - \bar{b}_i)(b_i - \bar{b}_i)^\top p(b_i \mid T_i, \delta_i, y_i; \hat{\theta}) \; db_i \\ H_i &= \left\{ -\frac{\partial^2 \log p(b | T_i, \delta_i, y_i; \hat{\theta})}{\partial b^\top \partial b} \Big|_{b = \hat{b}_i} \right\}^{-1} \end{aligned}\right.
$$

- *•* Bayesian estimation
	- \triangleright under the Bayesian paradigm both θ and $\{b_i, i=1,\ldots,n\}$ are regarded as parameters
- *•* Inference is based on the full posterior distribution

$$
p(\theta, b \mid T, \delta, y) = \frac{\prod_i p(T_i, \delta_i \mid b_i; \theta) p(y_i \mid b_i; \theta) p(b_i; \theta) p(\theta)}{\prod_i p(T_i, \delta_i, y_i)}
$$

$$
\propto \prod_{i=1}^n \Bigl\{ p(T_i, \delta_i \mid b_i; \theta) \; p(y_i \mid b_i; \theta) \; p(b_i; \theta) \Bigr\} \; p(\theta)
$$

- *•* No closed-form solutions for the integrals in the normalizing constant *⇒* **MCMC**
- For the standard joint model we have define thus far, the majority of the parameters can be updated using Gibbs sampling (or slice sampling)
	- *◃* when no close-form posterior conditionals are available, we can use the Metropolis-Hastings algorithm
- *•* To gain in efficiency, we can do block-updating for many of the parameters, i.e., *◃* fixed effects *β*
	- \triangleright random effects b_i
	- *◃* baseline covariates in the survival submodel *γ*

- *•* Good proposal distributions can be obtained from the separate fits of the two submodels
- *•* Not directly programmable in WinBUGS, INLA, etc., due to the integral in the definition of the survival function

$$
S_i(t | b_i; \theta) = \exp\left(-\int_0^t h_0(s; \theta) \exp\{\gamma^\top w_i + \alpha m_i(s)\} ds\right)
$$

extra steps required. . .

- *•* Inference then proceeds in the usual manner from the MCMC output, e.g.,
	- *◃* posterior means, variances, and standard errors
	- *◃* credible intervals
	- *◃* Bayes factors
	- *◃* DIC, CPO
	- \triangleright ...

• Example: To illustrate the virtues of joint modeling, we compare it with the standard time-dependent Cox model for the AIDS data

$$
\begin{cases}\ny_i(t) = m_i(t) + \varepsilon_i(t) \\
= \beta_0 + \beta_1 t + \beta_2 \{t \times \text{ddI}_i\} + b_{i0} + b_{i1}t + \varepsilon_i(t), \qquad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2), \\
h_i(t) = h_0(t) \exp{\gamma \text{ddI}_i + \alpha m_i(t)}\},\n\end{cases}
$$

where

 $\rhd h_0(t)$ is assumed piecewise-constant

• Clearly, there is a considerable effect of ignoring the measurement error, especially for the CD4 cell counts

- *•* A unit decrease in CD4¹*/*² , results in a
	- *◃* **Joint Model**: 1.3-fold increase in risk (95% CI: 1.24; 1.43)
	- *◃* **Time-Dependent Cox**: 1.2-fold increase in risk (95% CI: 1.16; 1.27)
- Which one to believe?
	- \triangleright a lot of theoretical and simulation work has shown that the Cox model underestimates the true association size of markers

R> Joint models are fitted using function jointModel() from package **JM**. This function accepts as main arguments a linear mixed model and a Cox PH model based on which it fits the corresponding joint model

```
lmeFit <- lme(CD4 ~ obstime + obstime:drug,
    random = \sim obstime | patient, data = aids)
```

```
coxFit \leq coxph(Surv(Time, death) \sim drug, data = aids.id, x = TRUE)
```

```
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
   method = "piecewise-PH-aGH")
```

```
summary(jointFit)
```


R> As before, the data frame given in $\text{Im}(c)$ should be in the long format, while the data frame given to coxph() should have one line per subject*[∗]*

 \rhd the ordering of the subjects needs to be the same

- R> In the call to $cosh()$ you need to set $x = TRUE$ (or model = TRUE) such that the design matrix used in the Cox model is returned in the object fit
- R> Argument timeVar specifies the time variable in the linear mixed model

[∗] Unless you want to include exogenous time-varying covariates or handle competing risks

R> Argument method specifies the type of relative risk model and the type of numerical integration algorithm – the syntax is as follows:

<baseline hazard>-<parameterization>-<numerical integration>

Available options are:

- *◃* "piecewise-PH-GH": PH model with piecewise-constant baseline hazard
- *◃* "spline-PH-GH": PH model with B-spline-approximated log baseline hazard
- *◃* "weibull-PH-GH": PH model with Weibull baseline hazard
- *◃* "weibull-AFT-GH": AFT model with Weibull baseline hazard
- *◃* "Cox-PH-GH": PH model with unspecified baseline hazard

GH stands for standard Gauss-Hermite; using aGH invokes the pseudo-adaptive Gauss-Hermite rule

R> Joint models under the Bayesian approach are fitted using function jointModelBayes() from package **JMbayes**. This function works in a very similar manner as function jointModel(), e.g.,

lmeFit <- lme(CD4 ~ obstime + obstime:drug, random = \sim obstime | patient, data = aids)

coxFit \leq coxph(Surv(Time, death) \sim drug, data = aids.id, x = TRUE)

```
jointFitBayes <- jointModelBayes(lmeFit, coxFit, timeVar = "obstime")
summary(jointFitBayes)
```


R> **JMbayes** is more flexible (in some respects):

▷ directly implements the MCMC

◃ allows for categorical longitudinal data as well

◃ allows for general transformation functions

◃ penalized B-splines for the baseline hazard function

 \triangleright ...

- R> In both packages methods are available for the majority of the standard generic functions $+$ extras
	- *◃* summary(), anova(), vcov(), logLik()
	- *◃* coef(), fixef(), ranef()
	- *◃* fitted(), residuals()
	- \rhd plot()
	- *◃* xtable() (you need to load package **xtable** first)

- So far we have attacked the problem from the survival point of view
- *•* However, often, we may be also interested on the longitudinal outcome
- *•* **Issue:** When patients experience the event, they dropout from the study \rhd a direct connection with the missing data field

Dropout must be taken into account when deriving inferences for the longitudinal outcome

- To show this connection more clearly
	- $\triangleright T_i^*$ i^* : true time-to-event
	- $\triangleright y_i^o$ $_i^o$: longitudinal measurements before T_i^* *i*
	- \triangleright y_i^m *i* : longitudinal measurements after *T ∗ i*
- **Important to realize** that the model we postulate for the longitudinal responses is for the complete vector $\{y^o_i\}$ $\{v_i^o, y_i^m\}$

◃ implicit assumptions about missingness

• Missing data mechanism:

$$
p(T^*_i \mid y^o_i, y^m_i) \ = \ \int p(T^*_i \mid b_i) \ p(b_i \mid y^o_i, y^m_i) \ db_i
$$

still depends on *y m* $\frac{m}{i}$, which corresponds to nonrandom dropout

> **Intuitive interpretation:** Patients who dropout show different longitudinal evolutions than patients who do not

- *•* Implications of nonrandom dropout
	- *◃* observed data do not constitute a random sample from the target population
- *•* This feature complicates the validation of the joint model's assumptions using standard residual plots
	- *◃* what is the problem: Residual plots may show systematic behavior due to dropout and not because of model misfit

- *•* What about censoring?
	- *◃* censoring also corresponds to a discontinuation of the data collection process for the longitudinal outcome
- *•* Likelihood-based inferences for joint models provide valid inferences when censoring is MAR
	- *◃* a patient relocates to another country (MCAR)
	- *◃* a patient is excluded from the study when her longitudinal response exceeds a prespecified threshold (MAR)
	- *◃* censoring depends on random effects (MNAR)

• Joint models belong to the class of *Shared Parameter Models*

$$
p(y_i^o, y_i^m, T_i^*) = \int p(y_i^o, y_i^m \mid b_i) \ p(T_i^* \mid b_i) \ p(b_i) db_i
$$

the association between the longitudinal and missingness processes is explained by the *shared* random effects *bⁱ*

- *•* The other two well-known frameworks for MNAR data are
	- *◃* Selection models

$$
p(y_i^o, y_i^m, T_i^*) = p(y_i^o, y_i^m) \ p(T_i^* \mid y_i^o, y_i^m)
$$

◃ Pattern mixture models:

$$
p(y_i^o, y_i^m, T_i^*) = p(y_i^o, y_i^m \mid T_i^*) \; p(T_i^*)
$$

• These two model families are primarily applied with discrete dropout times and cannot be easily extended to continuous time

- $|$ Example: $|$ In the AIDS data the association parameter α was highly significant, suggesting nonrandom dropout
- *•* A comparison between
	- *◃* linear mixed-effects model *⇒* MAR
	- *◃* joint model *⇒* MNAR
	- is warranted
- MAR assumes that missingness depends only on the observed data

$$
p(T^*_i \mid y^o_i, y^m_i) = p(T^*_i \mid y^o_i)
$$

• Minimal sensitivity in parameter estimates & standard errors

⇒ **Warning:** This does not mean that this is always the case!

Chapter 5

Extensions of Joint Models

• The standard joint model

$$
\begin{cases}\nh_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp{\{\gamma^\top w_i + \alpha m_i(t)\}}, \\
y_i(t) = m_i(t) + \varepsilon_i(t) \\
= x_i^\top(t)\beta + z_i^\top(t)b_i + \varepsilon_i(t),\n\end{cases}
$$

$$
\text{where } \mathcal{M}_i(t) = \{m_i(s), 0 \leq s < t\}
$$

• The standard joint model

$$
\begin{cases}\nh_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp{\{\gamma^\top w_i + \alpha m_i(t)\}}, \\
y_i(t) = m_i(t) + \varepsilon_i(t) \\
= x_i^\top(t)\beta + z_i^\top(t)b_i + \varepsilon_i(t),\n\end{cases}
$$

where $M_i(t) = \{m_i(s), 0 \le s < t\}$

Is this the only option? Is this the most optimal choice?

- Note: Inappropriate modeling of time-dependent covariates may result in surprising results
- Example: Cavender et al. (1992, J. Am. Coll. Cardiol.) conducted an analysis to test the effect of cigarette smoking on survival of patients who underwent coronary artery surgery
	- *◃* the estimated effect of current cigarette smoking was positive on survival although not significant (i.e., patients who smoked had higher probability of survival)
	- *◃* most of those who had died were smokers but many stopped smoking at the last follow-up before their death

We need to carefully consider the functional form of time-dependent covariates

• Let's see some possibilities. . .

• Lagged Effects: The hazard for an event at *t* is associated with the level of the marker at a previous time point:

$$
h_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp{\lbrace \gamma^\top w_i + \alpha m_i(t^c_+) \rbrace},
$$

where

$$
t_+^c = \max(t - c, 0)
$$

• Time-dependent Slopes: The hazard for an event at *t* is associated with both the current value and the slope of the trajectory at *t* (Ye et al., 2008, Biometrics):

$$
h_i(t | \mathcal{M}_i(t)) = h_0(t) \exp{\{\gamma^\top w_i + \alpha_1 m_i(t) + \alpha_2 m'_i(t)\}},
$$

where

$$
m_i'(t) = \frac{d}{dt} \{ x_i^\top(t) \beta + z_i^\top(t) b_i \}
$$

• Cumulative Effects: The hazard for an event at *t* is associated with the whole area under the trajectory up to *t*:

$$
h_i(t | \mathcal{M}_i(t)) = h_0(t) \exp\left\{\gamma^{\top} w_i + \alpha \int_0^t m_i(s) \ ds\right\}
$$

• Area under the longitudinal trajectory taken as a summary of $\mathcal{M}_i(t)$

• Weighted Cumulative Effects (convolution): The hazard for an event at *t* is associated with the area under the weighted trajectory up to *t*:

$$
h_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp \left\{ \gamma^\top w_i + \alpha \int_0^t \varpi(t-s) m_i(s) \ ds \right\},\,
$$

where $\varpi(\cdot)$ an appropriately chosen weight function, e.g.,

- *◃* Gaussian density
- *◃* Student's-*t* density
- \triangleright . . .

• Random Effects: The hazard for an event at *t* is associated only with the random effects of the longitudinal model:

$$
h_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp(\gamma^\top w_i + \alpha^\top b_i)
$$

- *•* Features:
	- *◃* avoids numerical integration for the survival function
	- *◃* interpretation of *α* more difficult, especially in high-dimensional random-effects settings

- Example: Sensitivity of inferences for the longitudinal process to the choice of the parameterization for the AIDS data
- *•* We use the same mixed model as before, i.e.,

$$
y_i(t) = m_i(t) + \varepsilon_i(t)
$$

$$
= \beta_0 + \beta_1 t + \beta_2 \{t \times \mathbf{d} \mathbf{d} \mathbf{I}_i\} + b_{i0} + b_{i1} t + \varepsilon_i(t)
$$

and the following four survival submodels

• Model I (current value)

$$
h_i(t) = h_0(t) \exp{\lbrace \gamma \mathbf{d} \mathbf{d} \mathbf{I}_i + \alpha_1 m_i(t) \rbrace}
$$

• Model II (current value + current slope)

$$
h_i(t) = h_0(t) \exp{\lbrace \gamma \mathbf{d} \mathbf{d} \mathbf{I}_i + \alpha_1 m_i(t) + \alpha_2 m'_i(t) \rbrace},
$$

where

 $\triangleright m'_i(t) = \beta_1 + \beta_2$ ddI $_i + b_{i1}$

• Model III (random slope)

$$
h_i(t) = h_0(t) \exp{\lbrace \gamma \mathbf{ddI}_i + \alpha_3 b_{i1} \rbrace}
$$

• Model IV (area)

$$
h_i(t) = h_0(t) \exp \Bigl\{\gamma \text{ddI}_i + \alpha_4 \int_0^t m_i(s) \ ds\Bigr\},
$$

where

$$
\triangleright \int_0^t m_i(s) \; ds = \beta_0 t + \frac{\beta_1}{2} t^2 + \frac{\beta_2}{2} \{ t^2 \times \mathsf{ddI}_i \} + b_{i0} t + \frac{b_{i1}}{2} t^2
$$

- *•* There are noticeable differences between the parameterizations
	- *◃* especially in the slope parameters
- *•* Therefore, a sensitivity analysis should not stop at the standard joint model parameterization but also consider alternative association structures

- R> Lagged effects can be fitted using the lag argument of jointModel(). For example, the following code fits a joint model for the PBC dataset with
	- *◃* random intercepts and random slopes for log serum bilirubin, and
	- *◃* a relative risk model with piecewise-constant baseline hazard and the *true* effect at the previous year

```
lmeFit \leq lme(log(serBilir) \sim year, random = \sim year | id, data = pbc2)
```

```
coxFit \leq coxph(Surv(years, status2) \sim 1, data = pbc2.id, x = TRUE)
```

```
jointFit <- jointModel(lmeFit, coxFit, timeVar = "year",
   method = "piecewise-PH-aGH", lag = 1)
```

```
summary(jointFit)
```


- R> For the time-dependent slopes and cumulative effects parameterizations, arguments parameterization and derivForm of jointModel() should be used
	- *◃* the first one just specifies whether we want to include a single or two terms involving *mi*(*t*) in the linear predictor of the survival submodel, options are
		- * parameterization = "value"
		- * parameterization = "slope"
		- * parameterization = "both"
	- *◃* the second one requires a few extra steps to specify we will see an example in the practical

- So far we have concentrated on a single continuous marker
- *•* But very often we may have several markers we wish to study, some of which could be categorical
- Example: In the PBC dataset we have used serum bilirubin as the most important marker, but during follow-up several other markers have been recorded
	- *◃* serum cholesterol (continuous)
	- *◃* edema (3 categories)
	- *◃* ascites (2 categories)

 \triangleright ...

We need to extend the basic joint model!

- *•* To handle multiple longitudinal markers of different types we use Generalized Linear Mixed Models
	- \triangleright We assume Y_{i1}, \ldots, Y_{iJ} for each subject, each one having a distribution in the exponential family, with expected value

$$
m_{ij}(t) = E(y_{ij}(t) | b_{ij}) = g_j^{-1} \{ x_{ij}^{\top}(t) \beta_j + z_{ij}^{\top}(t) b_{ij} \},
$$

with *g*(*·*) denoting a link function

◃ Correlation between the outcomes is built by assuming a multivariate normal distribution for the random effects

$$
b_i = (b_{i1}^\top, \ldots, b_{iJ}^\top)^\top \sim \mathcal{N}(0, D)
$$

• The expected value of each longitudinal marker is incorporated in the linear predictor of the survival submodel

$$
h_i(t) = h_0(t) \exp\left\{\gamma^\top w_i + \sum_{j=1}^J \alpha_j m_{ij}(t)\right\}
$$

- *•* Often multiple failure times are recorded
	- *◃* competing risks
	- *◃* recurrent events
- $|$ Example: $|$ In the PBC dataset \Rightarrow competing risks
	- *◃* Some patients received a liver transplantation
	- *◃* So far we have used the composite event, i.e. death or transplantation whatever comes first
	- *◃* When interest only is on one type of event, the other should be considered as a competing risk

• Joint models with competing risks:

$$
\begin{cases}\n y_i(t) = m_i(t) + \varepsilon_i(t) = x_i^\top(t)\beta + z_i^\top(t)b_i + \varepsilon_i(t), \\
 h_i^d(t) = h_0^d(t) \exp{\{\gamma_d^\top w_i + \alpha_d m_i(t)\}}, \\
 h_i^{tr}(t) = h_0^{tr}(t) \exp{\{\gamma_{tr}^\top w_i + \alpha_{tr} m_i(t)\}},\n\end{cases}
$$

where

 $\triangleright h_i^d$ $\frac{d}{i}(t)$ hazard function for death $\triangleright h_i^{tr}$ $\frac{tr}{dt}(t)$ hazard function for transplantation

- *•* Multiple Failure Times: recurrent events
- $|$ Example: $|$ In the PBC dataset \Rightarrow recurrent events
	- *◃* Patients showed irregular visiting patterns
	- *◃* So far, when we fitted the joint model we assumed that the visiting process is non-informative
	- *◃* If this assumption is violated, we should also model this process in order to obtain valid inferences

• Joint model with recurrent (visiting process) & terminal events

$$
\begin{cases}\ny_i(t) = m_i(t) + \varepsilon_i(t) = x_i^{\top}(t)\beta + z_i^{\top}(t)b_i + \varepsilon_i(t), \\
r_i(t) = r_0(t)\exp\{\gamma_r^{\top}w_{ri} + \alpha_r m_i(t) + \mathbf{v}_i\}, \\
h_i(t) = h_0(t)\exp\{\gamma_h^{\top}w_{hi} + \alpha_h m_i(t) + \zeta \mathbf{v}_i\},\n\end{cases}
$$

with

 \triangleright $r_i(t)$ hazard function for the recurrent events $\triangleright h_i(t)$ hazard function for the terminal event \rhd v_i frailty term accounting for the correlation in the recurrent events

- *•* Features of multivariate joint models
	- *◃* using CI is straightforward to extend joint models to multiple longitudinal outcomes of different types, and multiple failure times
	- *◃* computationally much more intensive due to requirement for high dimensional numerical integrations with respect to the random effects

- *•* Note: In the previous extensions of joint models, i.e.,
	- *◃* multiple longitudinal markers
	- *◃* multiple failure times

we used the default parameterization that includes the current value term $m_i(t)$ in the linear predictor of the survival submodel(s)

> Nonetheless, all the other parameterizations we have seen earlier are also applicable

Chapter 6

Dynamic Predictions, Discrimination & Calibration

- Nowadays there is great interest for prognostic models and their application to personalized medicine
- *•* Examples are numerous
	- *◃* cancer research, cardiovascular diseases, HIV research, . . .

Physicians are interested in accurate prognostic tools that will inform them about the future prospect of a patient in order to adjust medical care

- *•* We are interested in predicting survival probabilities for a new patient *j* that has provided a set of serum bilirubin measurements up to a specific time point *t*
- Example: We consider Patients 2 and 25 from the PBC dataset that have provided us with 9 and 12 serum bilirubin measurements, respectively
	- **▷ Dynamic Prediction** survival probabilities are dynamically updated as additional longitudinal information is recorded
- *•* We need to account for the endogenous nature of the marker
	- *◃* providing measurements up to time point *t ⇒* the patient was still alive at time *t*

• More formally, for a new subject *j* we have available measurements up to time point *t*

$$
\mathcal{Y}_j(t) = \{y_j(s), 0 \le s \le t\}
$$

and we are interested in

$$
\pi_j(u \mid t) = \Pr\bigl\{T_j^* \geq u \mid T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n\bigr\},\
$$

where

 \rhd where $u > t$, and

 $D \triangleright \mathcal{D}_n$ denotes the sample on which the joint model was fitted

- *•* We assume that the joint model has been fitted to the data at hand
- *•* Based on the fitted model we can estimate the conditional survival probabilities (Rizopoulos, 2011, Biometrics)

• $\pi_i(u \mid t)$ can be rewritten as

$$
\pi_j(u \mid t) \ = \ \int \frac{S_j\{u \mid \mathcal{M}_j(u, b_j, \theta); \theta\}}{S_j\{t \mid \mathcal{M}_j(t, b_j, \theta); \theta\}} \, p(b_j \mid T_j^* > t, \mathcal{Y}_j(t); \theta) \, db_j
$$

• A naive estimator for *πj*(*u | t*) can be constructed by plugging-in the MLEs and the Empirical Bayes estimates

$$
\tilde{\pi}_j(u \mid t) = \frac{S_j\{u \mid \mathcal{M}_j(u, \hat{b}_j, \hat{\theta}); \hat{\theta}\}}{S_j\{t \mid \mathcal{M}_j(t, \hat{b}_j, \hat{\theta}); \hat{\theta}\}}
$$

 \rhd this works relatively well in practice, but

◃ standard errors are difficult to compute

• It is convenient to proceed using a Bayesian formulation of the problem *⇒* π ^{*j*}(*u | t*) can be written as

$$
\Pr\left\{T_j^* \geq u \mid T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n\right\} = \int \Pr\left\{T_j^* \geq u \mid T_j^* > t, \mathcal{Y}_j(t); \theta\right\} p(\theta \mid \mathcal{D}_n) d\theta
$$

• We have already seen the first part of the integrand

$$
\Pr\left\{T_j^* \ge u \mid T_j^* > t, \mathcal{Y}_j(t); \theta\right\} =
$$
\n
$$
= \int \frac{S_j\{u \mid \mathcal{M}_j(u, b_j, \theta); \theta\}}{S_j\{t \mid \mathcal{M}_j(t, b_j, \theta); \theta\}} p(b_j \mid T_j^* > t, \mathcal{Y}_j(t); \theta) db_j
$$

• Provided that the sample size is sufficiently large, we can approximate the posterior of the parameters by

$$
\{\theta\mid \mathcal{D}_n\}\sim \mathcal{N}(\hat{\theta},\hat{\mathcal{H}}),
$$

where

 \triangleright $\hat{\theta}$ are the MLEs, and

 \triangleright $\hat{\mathcal{H}}$ their asymptotic covariance matrix

• A Monte Carlo estimate of *πj*(*u | t*) can be obtained using the following simulation scheme:

 $\mathsf{Step\ 1.}$ draw $\theta^{(\ell)} \sim \mathcal{N}(\hat{\theta}, \hat{\mathcal{H}})$

 $\textsf{Step 2.}$ draw $b_j^{(\ell)} \sim \{b_j \mid T_j^* > t, \mathcal{Y}_j(t), \theta^{(\ell)}\}$

Step 3. compute $\pi^{(\ell)}_i$ $S_j^{(\ell)}(u \mid t) = S_j\big\{u \mid \mathcal{M}_j(u, b_j^{(\ell)}, \theta^{(\ell)}); \theta^{(\ell)}\big\}\Big/S_j\big\{t \mid \mathcal{M}_j(t, b_j^{(\ell)}, \theta^{(\ell)}); \theta^{(\ell)}\big\}$

• Repeat Steps 1–3, *ℓ* = 1*, . . . , L* times, where *L* denotes the number of Monte Carlo samples

- Steps 1 and 3 are straightforward
- \bullet In Step 2 we need to sample from $\{b_j \mid T_j^* > t, \mathcal{Y}_j(t), \theta^{(\ell)}\}$, which is nonstandard
	- \triangleright as n_i increases, this posterior converges to a multivariate normal distribution (Rizopoulos et al., Biometrika, 2008)
	- \triangleright we use a Metropolis-Hastings algorithm with multivariate t proposals

- Example: Dynamic predictions of survival probabilities for Patients 2 & 25 from the PBC dataset: We fit the joint model
- *•* Longitudinal submodel
	- *◃* fixed effects: Linear & quadratic time, treatment and their interaction *◃* random effects: Intercept, linear & quadratic time effects
- *•* Survival submodel
	- \triangleright treatment effect $+$ *underlying* serum bilirubin level
	- *◃* piecewise-constant baseline hazard in 7 intervals

- Based on the fitted joint model we estimate $\pi_j(u \mid t)$ for Patients 2 and 25
- *•* We use 500 Monte Carlo samples, and we took as estimate

$$
\hat{\pi}_j(u \mid t) = \text{median}\{\pi_j^{(\ell)}(u \mid t), \ell = 1, \ldots, L\}
$$

and calculated a corresponding 95% pointwise CIs

R> Individualized predictions of survival probabilities are computed by function survfitJM() – for example, for Patient 2 from the PBC dataset we have

```
sfit \le survfitJM(jointFit, newdata = pbc2[pbc2$id == "2", ])
```
sfit

```
plot(sfit)
plot(sfit, include.y = TRUE)
```


- *•* In some occasions it may be also of interest to predict the longitudinal outcome
- We can proceed in the same manner as for the survival probabilities: We have available measurements up to time point *t*

$$
\mathcal{Y}_j(t) = \{y_j(s), 0 \le s \le t\}
$$

and we are interested in

$$
\omega_j(u \mid t) = E\big\{y_j(u) \mid T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n\big\}, \quad u > t
$$

6.3 Longitudinal Responses: Definitions*[∗]* **(cont'd)**

• To estimate $\omega_j(u \mid t)$ we can follow a similar approach as for $\pi_j(u \mid t)$ – Namely, $\omega_i(u \mid t)$ is written as:

$$
E\big\{y_j(u) | T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n\big\} = \int E\big\{y_j(u) | T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n; \theta\big\} p(\theta | \mathcal{D}_n) d\theta
$$

• With the first part of the integrand given by:

$$
E\{y_j(u) | T_j^* > t, \mathcal{Y}_j(t), \mathcal{D}_n; \theta\} =
$$

=
$$
\int \{x_j^{\top}(u)\beta + z_j^{\top}(u)b_j\} p(b_j | T_j^* > t, \mathcal{Y}_j(t); \theta) db_j
$$

 $\omega^{(\ell)}_i$

|Note: Prediction intervals can be easily computed by replacing Step 3 with a draw from:

 $\int\limits_j^\top (u) \beta^{(\ell)} + z_j^\top$

 $j^{\top}(u) b^{(\ell)}_j$

 $\left\{\begin{matrix}\ell\\j\end{matrix}\right\}, \quad \left[\sigma^2\right]^{(\ell)}$

Step 3. compute $\omega^{(\ell)}_i$ $y_j^{(\ell)}(u \mid t) = x_j^\top$ $\int\limits_j^\top (u) \beta^{(\ell)} + z_j^\top$ $j^{\top}(u) b^{(\ell)}_j$ *j*

 $y_j^{(\ell)}(u \mid t) \sim \mathcal{N} \left\{ x_j^\top \right\}$

 $\textsf{Step 2.}$ draw $b_j^{(\ell)} \sim \{b_j \mid T_j^* > t, \mathcal{Y}_j(t), \theta^{(\ell)}\}$

Step 1. draw
$$
\theta^{(\ell)} \sim \mathcal{N}(\hat{\theta}, \hat{\mathcal{H}})
$$

• A similar Monte Carlo simulation scheme:

6.3 Longitudinal Responses: Estimation*[∗]* **(cont'd)**

- Example: Dynamic predictions of serum bilirubin for Patients 2 & 25 from the PBC dataset: We fit the joint model
- *•* Longitudinal submodel
	- *◃* fixed effects: Linear & quadratic time, treatment and their interaction *◃* random effects: Intercept, linear & quadratic time effects
- *•* Survival submodel
	- \triangleright treatment effect $+$ *underlying* serum bilirubin level
	- *◃* piecewise-constant baseline hazard in 7 intervals

- Based on the fitted joint model we estimate $\omega_j(u \mid t)$ for Patients 2 and 25
- *•* Point estimates

$$
\hat{\omega}_j(u \mid t) = x_j^{\top}(u)\hat{\beta} + z_j^{\top}(u)\hat{b}_j,
$$

where $\hat{\beta}$: MLEs & \hat{b}_j : empirical Bayes estimates

• 95% pointwise Cls

◃ simulation scheme: 2.5% and 97.5% percentiles of 500 Monte Carlo samples of $\omega^{(\ell)}_i$ $j^{(\ell)}(u \mid t)$

R> Individualized predictions for the longitudinal outcome are computed by function predict() – for example, for Patient 2 from the PBC dataset we have function

```
lfit \leq predict(jointFit, newdata = pbc2[pbc2$id == "2", ],
    type = "Subject", interval = "conf", returnData = TRUE)
```
lfit

```
xyplot(pred + low + upp \tilde{ } year, data = lfit, type = "l",
    lty = c(1,2,2), col = c(2,1,1), lwd = 2)
```


R> Web interface using the **shiny** package

```
library("shiny")
```

```
runApp(file.path(.Library, "JMbayes/demo"))
```


• All previous predictions were based on the standard joint model

$$
\begin{cases}\nh_i(t \mid \mathcal{M}_i(t)) = h_0(t) \exp{\{\gamma^\top w_i + \alpha m_i(t)\}}, \\
y_i(t) = m_i(t) + \varepsilon_i(t) \\
= x_i^\top(t)\beta + z_i^\top(t)b_i + \varepsilon_i(t),\n\end{cases}
$$

$$
\text{ where } \mathcal{M}_i(t) = \{m_i(s), 0 \leq s < t\}
$$

- We have seen earlier that there are several alternative parameterizations (see Section 5.1)
- *•* Relevant questions:
	- *◃* Does the assumed parameterization affect predictions?
	- *◃* Which parameterization is the most optimal?
- **Example: We compare predictions for the longitudinal and survival outcomes under** different parameterizations for Patient 51 from the PBC study

- *•* Predictions based on five joint models for the PBC dataset
	- *◃* the same longitudinal submodel as before, and

◃ relative risk submodels:

 $h_i(t) = h_0(t) \exp{\{\gamma \mathbf{D} - \mathbf{pnc}_i + \alpha_1 m_i(t)\}},$

$$
h_i(t) \ = \ h_0(t) \exp\{\gamma \text{D-pnc}_i + \alpha_2 m_i'(t)\},
$$

$$
h_i(t) \ = \ h_0(t) \exp\{\gamma \text{D-pnc}_i + \alpha_1 m_i(t) + \alpha_2 m_i'(t)\},
$$

$$
h_i(t) \ = \ h_0(t) \exp\Bigl\{\gamma{\bf D}\text{-}{\bf pn}{\bf c}_i + \alpha_3 \int_0^t m_i(s)ds\Bigr\},
$$

$$
h_i(t) \ = \ h_0(t) \exp\Bigl\{\gamma{\bf D}\text{-}{\bf pn}{\bf c}_i + \alpha_4 \int_0^t \phi(t-s) m_i(s) ds\Bigr\},
$$

where *ϕ*(*·*) standard normal pdf

Longitudinal Outcome

Predicted log serum bilirubin

Survival Outcome

- *•* The chosen parameterization can influence the derived predictions \rhd especially for the survival outcome
- *•* My current work: How to optimally choose parameterization? *◃* per subject (personalized medicine)
- *•* Quite promising results from the Bayesian approach using Bayesian Model Averaging techniques
	- *◃* it can be done with package **JMbayes**,
	- \triangleright it falls a bit outside the scope of this course, but
	- **▷ I can provide information if interested...**

Chapter 7 Closing

• **When we need joint models for longitudinal and survival outcomes?**

◃ to handle endogenous time-varying covariates in a survival analysis context *◃* to account for nonrandom dropout in a longitudinal data analysis context

• **How joint models work?**

- *◃* a mixed model for the longitudinal outcome
- *◃* a relative risk model for the event process
- *◃* explain interrelationships with shared random effects

• **Where to pay attention when defining joint models?**

- *◃* model flexibly the subject-specific evolutions for the longitudinal outcome
- \triangleright use parametric but flexible models for the baseline hazard function
- *◃* consider how to model the association structure between the two processes *⇒* Parameterization

• **Extensions**

- *◃* under the full conditional independence assumption we can easily extend the basic joint model
- \rhd multiple longitudinal outcomes and/or multiple failure times
- \triangleright though more computationally intensive

• **Individualized predictions**

- *◃* joint models can provide subject-specific predictions for the longitudinal and survival outcomes
- *◃* these are dynamically updated as extra information is recorded for the subjects
- *◃ ⇒* joint models constitute an excellent tool for personalized medicine

• **What we did not cover**

- *◃* assessment of predictive performance
- **▷ diagnostics for joint models using residuals**

 \triangleright ...

The End!

- *•* Andrinopoulou, E.R., Rizopoulos, D., Takkenberg, J. and Lesaffre, E. (2014). Joint modeling of two longitudinal outcomes and competing risk data. *Statistics in Medicine*, to appear.
- *•* Brown, E. and Ibrahim, J. (2003). A Bayesian semiparametric joint hierarchical model for longitudinal and survival data. *Biometrics* **59**, 221–228.
- *•* Brown, E. Ibrahim, J. and DeGruttola, V. (2005). A flexible B-spline model for multiple longitudinal biomarkers and survival. *Biometrics* **61**, 64–73.
- *•* Chi, Y.-Y. and Ibrahim, J. (2006). Joint models for multivariate longitudinal and multivariate survival data. *Biometrics* **62**, 432–445.
- *•* DeGruttola, V. and Tu, X. (1994). Modeling progression of CD-4 lymphocyte count and its relationship to survival time. *Biometrics* **50**, 1003–1014.
- *•* Elashoff, R., Li, G. and Li, N. (2008). A joint model for longitudinal measurements and survival data in the presence of multiple failure types. *Biometrics* **64**, 762–771.

- *•* Faucett, C. and Thomas, D. (1996). Simultaneously modelling censored survival data and repeatedly measured covariates: A Gibbs sampling approach. *Statistics in Medicine* **15**, 1663–1685.
- *•* Gerds, T. and Schumacher, M. (2006). Consistent estimation of the expected Brier score in general survival models with right-censored event times. *Biometrical Journal* **48**, 1029–1040.
- *•* Heagerty, P. and Zheng, Y. (2005). Survival model predictive accuracy and ROC curves. *Biometrics* **61**, 92–105.
- *•* Henderson, R., Diggle, P. and Dobson, A. (2000). Joint modelling of longitudinal measurements and event time data. *Biostatistics* **1**, 465–480.
- *•* Henderson, R., Diggle, P. and Dobson, A. (2002). Identification and efficacy of longitudinal markers for survival. *Biostatistics* **3**, 33–50.
- *•* Hsieh, F., Tseng, Y.-K. and Wang, J.-L. (2006). Joint modeling of survival and longitudinal data: Likelihood approach revisited. *Biometrics* **62**, 1037–1043.

- *•* Lin, H., Turnbull, B., McCulloch, C. and Slate, E. (2002). Latent class models for joint analysis of longitudinal biomarker and event process: Application to longitudinal prostate-specific antigen readings and prostate cancer. *Journal of the American Statistical Association* **97**, 53–65.
- *•* Liu, L. and Huang, X. (2009). Joint analysis of correlated repeated measures and recurrent events processes in the presence of death, with application to a study on acquired immune deficiency syndrome. *Journal of the Royal Statistical Society, Series C* **58**, 65–81.
- *•* Proust-Lima, C., Joly, P., Dartigues, J. and Jacqmin-Gadda, H. (2009). Joint modelling of multivariate longitudinal outcomes and a time-to-event: A nonlinear latent class approach. *Computational Statistics and Data Analysis* **53**, 1142–1154.
- *•* Proust-Lima, C. and Taylor, J. (2009). Development and validation of a dynamic prognostic tool for prostate cancer recurrence using repeated measures of posttreatment PSA: A joint modeling approach. *Biostatistics* **10**, 535–549.
- *•* Rizopoulos, D. (2012). Fast fitting of joint models for longitudinal and event time data using a pseudo-adaptive Gaussian quadrature rule. *Computational Statistics and Data Analysis* **56**, 491–501.
- *•* Rizopoulos, D. (2011). Dynamic predictions and prospective accuracy in joint models for longitudinal and time-to-event data. *Biometrics* **67**, 819–829.

- *•* Rizopoulos, D. (2010). JM: An R package for the joint modelling of longitudinal and time-to-event data. *Journal of Statistical Software* **35** (9), 1–33.
- *•* Rizopoulos, D. and Ghosh, P. (2011). A Bayesian semiparametric multivariate joint model for multiple longitudinal outcomes and a time-to-event. *Statistics in Medicine* **30**, 1366–1380.
- *•* Rizopoulos, D., Hatfield, L.A., Carlin, B.P. and Takkenberg, J.J.M. (2014). Combining dynamic predictions from joint models for longitudinal and time-to-event data using Bayesian model averaging. *Journal of the American Statistical Association* **109**, 1385–1397.
- *•* Rizopoulos, D., Murawska, M., Andrinopoulou, E.-R., Molenberghs, G., Takkenberg, J. and Lesaffre, E. (2013). Dynamic predictions with time-dependent covariates in survival analysis using joint modeling and landmarking. *Submitted*.
- *•* Rizopoulos, D. and Lesaffre, E. (2014). Introduction to the special issue on joint modelling techniques. *Statistical Methods in Medical Research* **23**, 3–10.
- *•* Rizopoulos, D., Verbeke, G. and Lesaffre, E. (2009). Fully exponential Laplace approximation for the joint modelling of survival and longitudinal data. *Journal of the Royal Statistical Society, Series B* **71**, 637–654.

- *•* Rizopoulos, D., Verbeke, G., Lesaffre, E. and Vanrenterghem, Y. (2008). A two-part joint model for the analysis of survival and longitudinal binary data with excess zeros. *Biometrics* **64**, 611–619.
- *•* Rizopoulos, D., Verbeke, G. and Molenberghs, G. (2010). Multiple-imputation-based residuals and diagnostic plots for joint models of longitudinal and survival outcomes. *Biometrics* **66**, 20–29.
- *•* Rizopoulos, D., Verbeke, G. and Molenberghs, G. (2008). Shared parameter models under random effects misspecification. *Biometrika* **95**, 63–74.
- *•* Rubin, D. (1976). Inference and missing data. *Biometrika* **63**, 581–592.
- *•* Song, X., Davidian, M. and Tsiatis, A. (2002). A semiparametric likelihood approach to joint modeling of longitudinal and time-to-event data. *Biometrics* **58**, 742–753.
- *•* Taylor, J., Park, Y., Ankerst, D., Proust-Lima, C., Williams, S., Kestin, L., Bae, K., Pickles, T., and Sandler, H. (2013). Real-time individual predictions of prostate cancer recurrence using joint models. *Biometrics*, **69**, 206–213.
- *•* Tseng, Y.-K., Hsieh, F. and Wang, J.-L. (2005). Joint modelling of accelerated failure time and longitudinal data. *Biometrika* **92**, 587–603.

- *•* Tsiatis, A. and Davidian, M. (2001). A semiparametric estimator for the proportional hazards model with longitudinal covariates measured with error. *Biometrika* **88**, 447–458.
- *•* Tsiatis, A. and Davidian, M. (2004). Joint modeling of longitudinal and time-to-event data: An overview. *Statistica Sinica* **14**, 809–834.
- *•* Tsiatis, A., DeGruttola, V., and Wulfsohn, M. (1995). Modeling the relationship of survival to longitudinal data measured with error: Applications to survival and CD4 counts in patients with AIDS. *Journal of the American Statistical Association* **90**, 27–37.
- Viviani, S., Alfó, M. and Rizopoulos, D. (2014). Generalized linear mixed joint model for longitudinal and survival outcomes. *Statistics and Computing*, **24**, 417–427.
- Viviani, S., Rizopoulos, D. and Alfó, M. (2014). Local sensitivity of shared parameter models to nonignorability of dropout. *Statistical Modelling* **14**, 205–228.
- *•* Wang, Y. and Taylor, J. (2001). Jointly modeling longitudinal and event time data with application to acquired immunodeficiency syndrome. *Journal of the American Statistical Association* **96**, 895–905.

- *•* Wu, M. and Bailey, K. (1988). Analysing changes in the presence of informative right censoring caused by death and withdrawal. *Statistics in Medicine* **7**, 337–346.
- *•* Wu, M. and Bailey, K. (1989). Estimation and comparison of changes in the presence of informative right censoring: conditional linear model. *Biometrics* **45**, 939–955.
- *•* Wu, M. and Carroll, R. (1988). Estimation and comparison of changes in the presence of informative right censoring by modeling the censoring process. *Biometrics* **44**, 175–188.
- *•* Wulfsohn, M. and Tsiatis, A. (1997). A joint model for survival and longitudinal data measured with error. *Biometrics* **53**, 330–339.
- *•* Xu, C., Baines, P. and Wang, J.-L. (2014). Standard error estimation using the EM algorithm for the joint modeling of survival and longitudinal data. *Biostatistics*, to appear.
- *•* Xu, J. and Zeger, S. (2001). Joint analysis of longitudinal data comprising repeated measures and times to events. *Applied Statistics* **50**, 375–387.

- *•* Ye, W., Lin, X., and Taylor, J. (2008). Semiparametric modeling of longitudinal measurements and time-to-event data a two stage regression calibration approach. *Biometrics* **64**, 1238–1246.
- *•* Yu, M., Law, N., Taylor, J., and Sandler, H. (2004). Joint longitudinal-survival-cure models and their application to prostate cancer. *Statistica Sinica* **14**, 835–862.
- *•* Yu, M., Taylor, J. and Sandler, H. (2008). Individualized prediction in prostate cancer studies using a joint longitudinal-survival-cure model. *Journal of the American Statistical Association* **108**, 178–187.
- *•* Zeng, D. and Cai, J. (2005). Asymptotic results for maximum likelihood estimators in joint analysis of repeated measurements and survival time. *The Annals of Statistics* **33**, 2132–2163.
- *•* Zheng, Y. and Heagerty, P. (2007). Prospective accuracy for longitudinal markers. *Biometrics* **63**, 332–341.

- *•* Andrinopoulou, E.R., Rizopoulos, D., Jin, R., Bogers, A., Lesaffre, E. and Takkenberg, J. (2012). An introduction to mixed models and joint modeling: Analysis of valve function over time. *Annals of Thoracic Surgery* **93**, 1765–1772.
- *•* Andrinopoulou, E.R., Rizopoulos, D., Geleijnse, M., Lesaffre, E., Bogers, A. and Takkenberg, J. (2015). Dynamic prediction of outcome for patients with severe aortic stenosis: Application of joint models for longitudinal and time-to-event data. *BMC Cardiovascular Disorders*, to appear.
- *•* Daher Abdi, D.Z., Essig, M., Rizopoulos, D., Le Meur, Y., Premaud, A., Woillard, J.-B., Rerolle, J.-P., Marquet, P. and Rousseau, A. (2013). Impact of longitudinal exposure to mycophenolic acid on acute rejection in renal-transplant recipients using a joint modeling approach. *Pharmacological Research* **72**, 52–60.
- *•* Ibrahim, J., Chu, H. and Chen, L.-M. (2010). Basic concepts and methods for joint models of longitudinal and survival data. *Journal of Clinical Oncology* **28**, 2796–2801.
- *•* Nunez, J., Nunez, E., Rizopoulos, D., Minana, G., Bodi, V., Bondanza, L., Husser, O., Merlos, P., Santas, E., Pascual-Figal, D.,; Chorro, F. and Sanchis, J. (2014). Red blood cell distribution width is longitudinally associated with mortality and incident anemia in heart failure patients. *Circulation Journal* **78**, 410—418.
- *•* Rizopoulos, D. and Takkenberg, J. (2014). Tools & Techniques: Dealing with time-varying covariates in survival analysis - joint models versus Cox models. *EuroIntervention* **10**, 285–288.

- *•* Thabut, G., Christie, J., Mal, H., Fournier, M., Brugiere, O., Leseche, G., Castier, Y. and Rizopoulos, D. (2013). Survival benefit of lung transplant for cystic fibrosis since lung-allocation-score implementation. *American Journal of Respiratory and Critical Care Medicine* **187**, 1335–1340.
- *•* van der Linde, D., Roos-Hesselink, J., Rizopoulos, D., Heuvelman, H., Budts, W., van Dijk, A., Witsenburg, M., Yap, S., Bogers, A., Oxenius, A., Silversides, C., Oechslin, E. and Takkenberg, J. (2013). Surgical outcome of discrete subaortic stenosis in adults: A multicenter study. *Circulation* **127**, 1184–1191.
- *•* van der Linde, D., Takkenberg, J., Rizopoulos, D., Heuvelman, H., Budts, W., van Dijk, A., Witsenburg, M., Yap, S., Bogers, A., Oxenius, A., Silversides, C., Oechslin, E. and Roos-Hesselink, J. (2013). Natural history of discrete subaortic stenosis in adults: A multicenter study. *European Heart Journal* **34**, 1548–1556.

Practicals

- *•* We will fit a simple joint model to the PBC dataset
- Start R and load package **JM**, using library (JM)
- *•* The longitudinal (long format) and survival information for the PBC patients can be found in data frames pbc2 and pbc2.id. The variables that we will need are:

◃ pbc2

- * id: patient id number
- * serBilir: serum bilirubin
- * year: follow-up times in years

◃ pbc2.id

- * years: observed event times in years
- * status: 'alive', 'transplanted', 'dead'
- * drug: treatment indicator

• T1: Fit the linear mixed effects model for log serum bilirubin using function $\text{Im}(\epsilon)$, assuming simple linear evolutions in time for each subject, i.e., a simple random-intercepts and random-slopes structure and different average evolutions per treatment group (see pp. 31–35)

$$
y_i(t) = \beta_0 + \beta_1 t + \beta_2 \{ \mathsf{D-penic}_i \times t \} + b_{i0} + b_{i1}t + \varepsilon_i(t)
$$

• T2: Create the indicator for the composite event (i.e., 'alive' = 0, 'transplanted' or 'dead' $= 1$) using the code

pbc2.id\$status2 <- as.numeric(pbc2.id\$status != "alive")

- T3: Fit the Cox PH model using coxph() that includes only treatment as baseline covariate, remember to set $x = TRUE$ (see pp. 55–56)
- *•* We want to fit the joint model

$$
\left\{ \begin{array}{l} y_i(t) \ = \ m_i(t) + \varepsilon_i(t) \\ \\ \qquad = \ \beta_0 + \beta_1 t + \beta_2 \{ \texttt{D-penic}_i \times t \} + b_{i0} + b_{i1} t + \varepsilon_i(t), \quad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2), \\ \\ h_i(t) \ = \ h_0(t) \exp\{\gamma \texttt{D-penic}_i + \alpha m_i(t) \}, \end{array} \right.
$$

• T4: Fit this joint model based on the fitted linear mixed and Cox models using function jointModel() (see pp. 93-95)

◃ with piecewise-constant baseline hazard & the (pseudo) adaptive GH rule

- T5: Use the summary () method to obtain a detailed output of the fitted joint model – interpret the results
- T6: Produce 95% confidence intervals for the parameters in the longitudinal submodel, and for the hazard ratios in the survival submodel using function $\text{confint}()$ (the parm argument of confint() can take as values "all" (default), "Longitudinal" and "Event")

- *•* This model assumes that the strength of the association between the level of serum bilirubin and the risk for the composite event is the same in the the two treatment groups
- *•* To relax this additivity assumption we will add the interaction effect between serum bilirubin and treatment

$$
\left\{ \begin{array}{l} y_i(t) \ = \ m_i(t) + \varepsilon_i(t) \\ \\ \qquad = \ \beta_0 + \beta_1 t + \beta_2 \{ \texttt{D-penic}_i \times t \} + b_{i0} + b_{i1} t + \varepsilon_i(t), \quad \varepsilon_i(t) \sim \mathcal{N}(0, \sigma^2), \\ \\ h_i(t) \ = \ h_0(t) \exp\bigl[\gamma \texttt{D-penic}_i + \alpha_1 m_i(t) + \alpha_2 \{ \texttt{D-penic}_i \times m_i(t) \} \bigr], \end{array} \right.
$$

- *•* To fit this model with package **JM** we need to define the interFact argument of jointModel(). This should be a named list with two elements:
	- *◃* value: a formula with the factors for which we wish to calculate the interaction terms
	- *◃* data: the data frame used to fit the Cox model
- T7: Define this list and fit the corresponding joint model. Use the summary () method to obtained a detailed output and interpret the results

• Based on the fitted joint model we can test for three treatment effects, namely *◃* in the longitudinal process:

$$
H_0: \beta_2=0
$$

▷ in the survival process:

$$
H_0: \gamma = \alpha_2 = 0
$$

◃ in the joint process:

$$
H_0: \beta_2 = \gamma = \alpha_2 = 0
$$

- We would like test these hypotheses using likelihood ratio tests
- T8: Fit the three joint models under the corresponding H_0 , and use function anova() to perform the LRTs (this function accepts as a first argument the joint model under the null, and as second the joint model under the alternative)

- *•* We will work with the Liver Cirrhosis dataset
	- *◃* a placebo-controlled randomized trial on 488 liver cirrhosis patients
- Start R and load package **JM**, using library (JM)
- *•* The longitudinal (long format) and survival information for the liver cirrhosis patients can be found in data frames prothro and prothros, respectively. The variables that we will need are:

◃ prothro

- * id: patient id number
- * pro: prothrobin measurements
- * time: follow-up times in years
- * treat: randomized treatment

◃ prothros

- * Time: observed event times in years
- * death: event indicator with $0 = '$ alive', and $1 = '$ dead'
- * treat: randomized treatment

- *•* We will fit the following joint model to the Liver Cirrhosis dataset
	- *◃* longitudinal submodel: linear subject-specific random slopes for prothrobin levels allowing for different average evolutions in the two treatment groups

$$
y_i(t) = m_i(t) + \varepsilon_i(t)
$$

$$
m_i(t) = \beta_0 + \beta_1 t + \beta_2 \{ \text{Trt}_i \times t \} + b_{i0} + b_{i1} t
$$

◃ survival submodel: treatment effect & *true* effect of prothrobin

$$
h_i(t) = h_0(t) \exp{\{\gamma \text{Tr} \mathbf{t}_i + \alpha m_i(t)\}}
$$

 $h_0(t)$ taken piecewise-constant

- $\boxed{71}$: Fit the linear mixed model using $\boxed{1me}$ (), the Cox model using coxph(), and the corresponding joint model using jointModel()
- We are interested in producing predictions of survival probabilities for Patient 155
- **T2:** Extract the data of Patient 155 using the code

dataP155 <- prothro[prothro\$id == 155,]

- T3: Using the first measurement of Patient 155, and the fitted joint model calculate his conditional survival probabilities using function survfitJM() and plot it using the plot method (see p. 153)
- T4: Repeat the same procedure by including each time the next measurement of Patient 155 and see how his survival probabilities evolve dynamically in time as extra prothrobin measurements are recorded
	- *◃* check arguments conf.int and fill.area of the plot() method for including the 95% confidence intervals

- T5: Similarly, produce predictions for future longitudinal responses of Patient 155 using the predict() method for fitted joint models (see p. 160)
	- *◃* first using only the first measurement,
	- *◃* and following update the predictions after each new longitudinal measurement has been recorded